

A Single Mouse Trial Platform for Evaluation of Novel Agents in Acute Lymphoblastic Leukemia by the Pediatric Preclinical Testing Consortium

Richard B. Lock^{1*}, Kathryn Evans¹, Tara Pritchard¹, Cara Toscan¹, Chelsea Mayoh¹, Beverly Teicher², Raushan T. Kurmasheva³, Peter J. Houghton³ and Malcolm Smith².
¹Children's Cancer Institute, Sydney, Australia; ²National Cancer Institute, Bethesda, MD; ³Greehey Children's Cancer Research Institute, San Antonio, TX.



1. Introduction

- The outcome for several high-risk subtypes of pediatric acute lymphoblastic leukemia (ALL) is extremely poor.
- Selecting the most active agents for clinical evaluation is critical as there are relatively few patients eligible for clinical trials.
- Conventional preclinical testing of novel agents is not sufficiently resourced to be able to encompass the vast heterogeneity between and within pediatric ALL subtypes.
- New approaches to preclinical testing in pediatric ALL are required.
- A single mouse trial (SMT) platform using a large panel of pediatric ALL patient-derived xenografts (PDXs) allows:
 - preclinical assessment of novel agents on an almost clinical trial scale;
 - the broad heterogeneity of pediatric ALL to be approximated within a single experiment;
 - biomarker discovery and validation by using molecularly annotated PDXs.

2. Study Methods

- Agent administration:**
- Topotecan (Tpt), 0.6mg/kg IP daily × 5 × 2 weeks, repeated at 21 days.
 Birinapant (Bpt), 15mg/kg IP every 3 days × 5.
- Study design and analysis:**
- 80 pediatric ALL PDXs broadly representative of all pediatric ALL subtypes were molecularly annotated by RNA-seq, exome-seq and DNA copy number analysis.
 - 2 NSG mice/PDX were inoculated via tail vein injection and treatment began when the percentage of human CD45⁺ cells (%huCD45⁺) in the murine peripheral blood (PB) reached ≥ 1%.
 - An event was defined as ≥ 25% huCD45⁺ cells in PB, or when the mouse exhibited leukemia-related morbidity associated with high-level leukemic infiltration (≥ 50% huCD45⁺) of at least 2 major organs.
 - The Kaplan-Meier method was used to determine event-free survival (EFS) between control and treated groups.
 - Treatment response was monitored using Objective Response Measures (ORM) modeled after stringent clinical criteria, which was assessed at Day 42 post treatment initiation (Houghton et al., 2007);
 - PD1 = progressive disease 1, %huCD45⁺ in PB never drops below 1% and event is reached by Day 14.
 - PD2 = progressive disease 2, %huCD45⁺ in PB never drops below 1% and event is reached after Day 14, but before Day 42.
 - SD = stable disease, %huCD45⁺ in PB never drops below 1% and event is not reached by Day 42.
 - PR = partial response, %huCD45⁺ in PB < 1% for 1 week and event is not reached by Day 42.
 - CR = complete response, %huCD45⁺ in PB < 1% for 2 consecutive weeks and event is not reached by Day 42.
 - MCR = maintained complete response, %huCD45⁺ in PB < 1% for at least 3 consecutive weeks after treatment completion and event is not reached by Day 42.
 - Waterfall plots represent the ratio of the minimal %huCD45⁺ in the PB at any point after treatment initiation relative to the %huCD45⁺ at Day 0.
 - PDX authenticity was verified using a 60-allele SNP array at both inoculation and at event (El-Hoss et al., 2016).

3. Results

Retrospective Analysis of Single Mouse Data

- Retrospective analysis of >700 randomly selected mice from agents previously tested by our group (Jones et al., 2016) showed that the single mouse results predicted the overall group response from conventional testing 73.9% of the time (Figure 1).
- This increased to 85.8% if a deviation of ± one objective response measure was allowed (Figure 1).
- Historically, two major sources of exclusion from an experiment were bad cell source (PDX stocks contaminated with mouse thymoma) and tumor (mouse origin) (Figure 2).
- Elimination of bad cell source and mouse tumors by using the NSG mouse strain could increase the single mouse prediction of overall group response to > 90%.

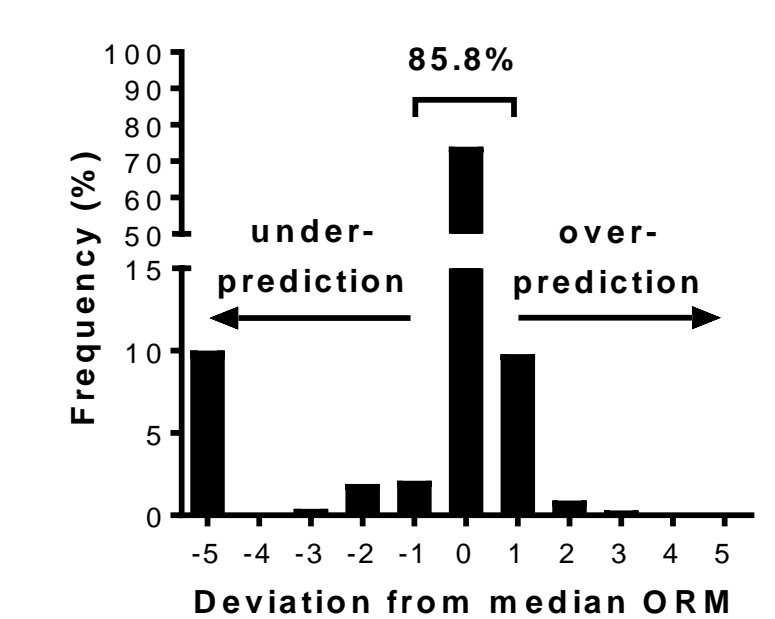


Figure 1. Concordance within conventional testing. The ORM of randomly selected mice was compared to the median ORM of their cohort. Single mice scored a 0 if their ORM matched their cohort median ORM. Single mice scored a +1 or -1 if their ORM was one category better or worse (respectively) than their cohort median ORM. Single mice that were excluded were assigned a -5 deviation score.

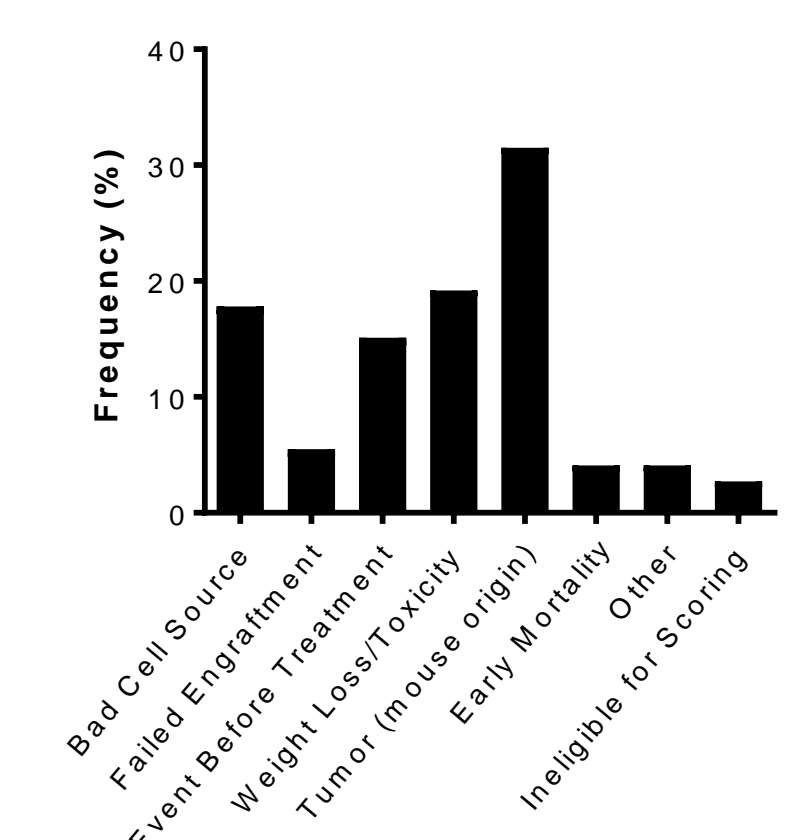


Figure 2. Frequency and source of exclusion of single mice. Single mice that were assigned a -5 deviation from their cohort median ORM in Figure 1.

SMT Pilot Study with Birinapant and Topotecan

- SMT results were achieved for 72 (90.0%) and 71 (88.8%) of the intended 80 mice for birinapant and topotecan, respectively.
- Waterfall plots revealed that 30/72 (41.7%) and 60/71 (84.5%) of PDXs achieved regressions in response to birinapant and topotecan, respectively (Figure 3). Distinctive activity profiles were identified for each agent.
- Comparing historical ORMs from conventional drug testing performed by the PPTC with SMT results showed high concordance for both birinapant (r = 0.804, p < 0.0001, n = 17) and topotecan (r = 0.904, p = 0.0143, n = 7) (Table 1).

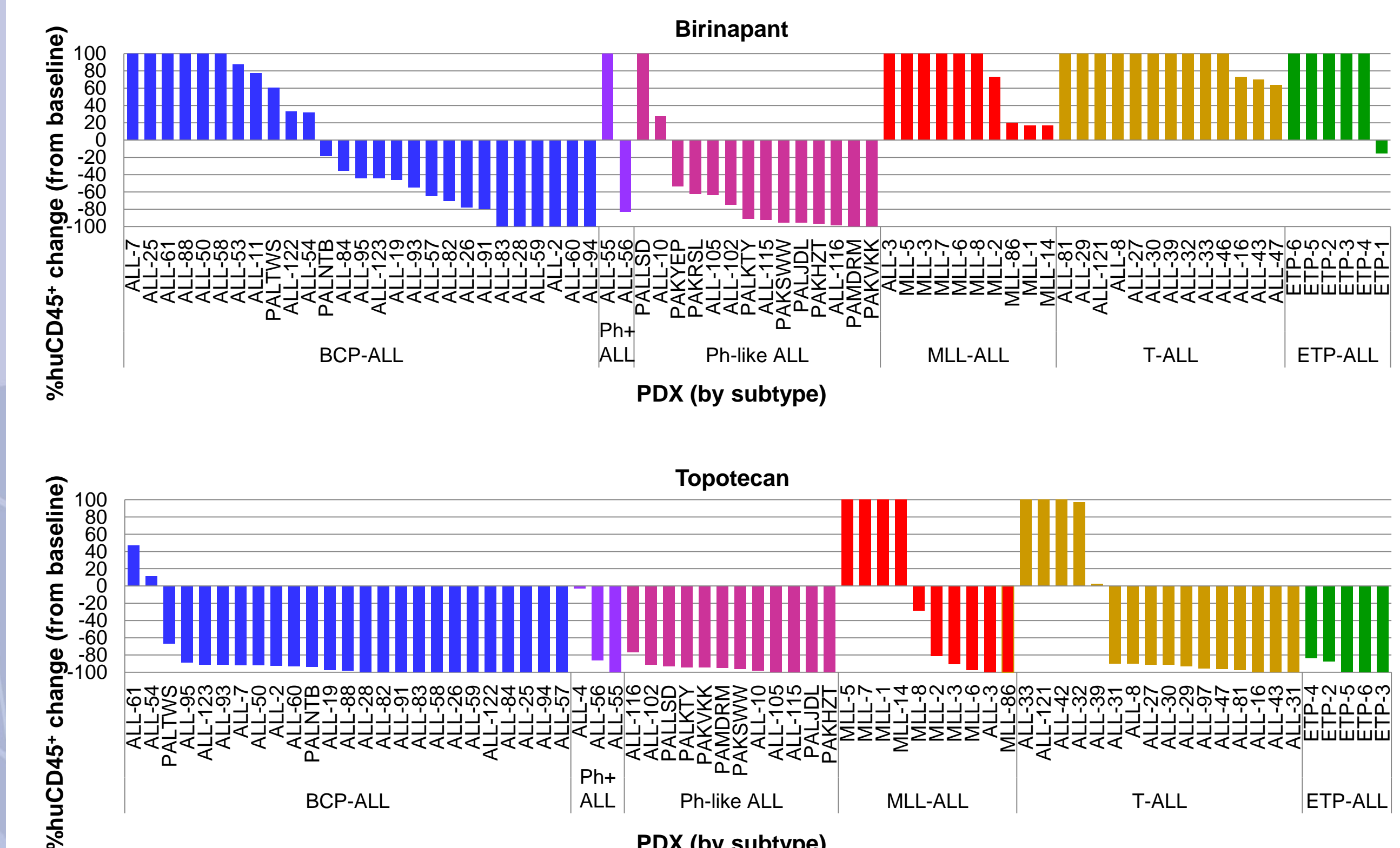


Figure 3. Waterfall plots depicting the maximum decrease from baseline (Day 0) levels of huCD45⁺ cells in PB. Birinapant treated (top) and topotecan treated (bottom). Each bar represents a single mouse. The graph is capped at ± 100%

Table 1. Comparison of SMT and historical data. Median ORM of conventional testing versus ORM of a single mouse from SMT for available PDXs for birinapant (top) and topotecan (bottom).

	Birinapant	
	Conventional 30 mg/kg	SMT 15 mg/kg
ALL-2	MCR	MCR
ALL-7	PD1	PD1
ALL-19	SD	PD1
ALL-50	PD2	PD1
ALL-54	PR	PD2
ALL-55	PD2	PD2
PALNTB	CR	PD2
PAKHZT	MCR	CR
PAKRSL	CR	PD2
PALLSD	CR	PD2
PAMDRM	MCR	MCR
PALJDL	CR	PR
ALL-10	CR	PD2
MLL-5	PD2	PD2
MLL-6	PD2	PD2
MLL-7	PD2	PD1
MLL-14	CR	PD2
# Mice	~304	17
	r = 0.804	p < 0.0001

	Topotecan	
	Conventional 0.6mg/kg	SMT 0.6mg/kg
ALL-2	MCR	MCR
ALL-3	CR	MCR
ALL-4	PD2	PD2
ALL-7	PR	CR
ALL-8	CR	CR
ALL-16	MCR	MCR
ALL-19	PR	PR
# Mice	~128	7
	r = 0.904	p = 0.0143

3. Results (continued)

Gene Expression Signatures Associated with In Vivo Response

- Analysis of divergent responses observed within the BCP-ALL subtype to birinapant and the within the MLL-ALL subtype to topotecan, revealed unique gene expression signatures that distinguished between *in vivo* response (Figure 4).

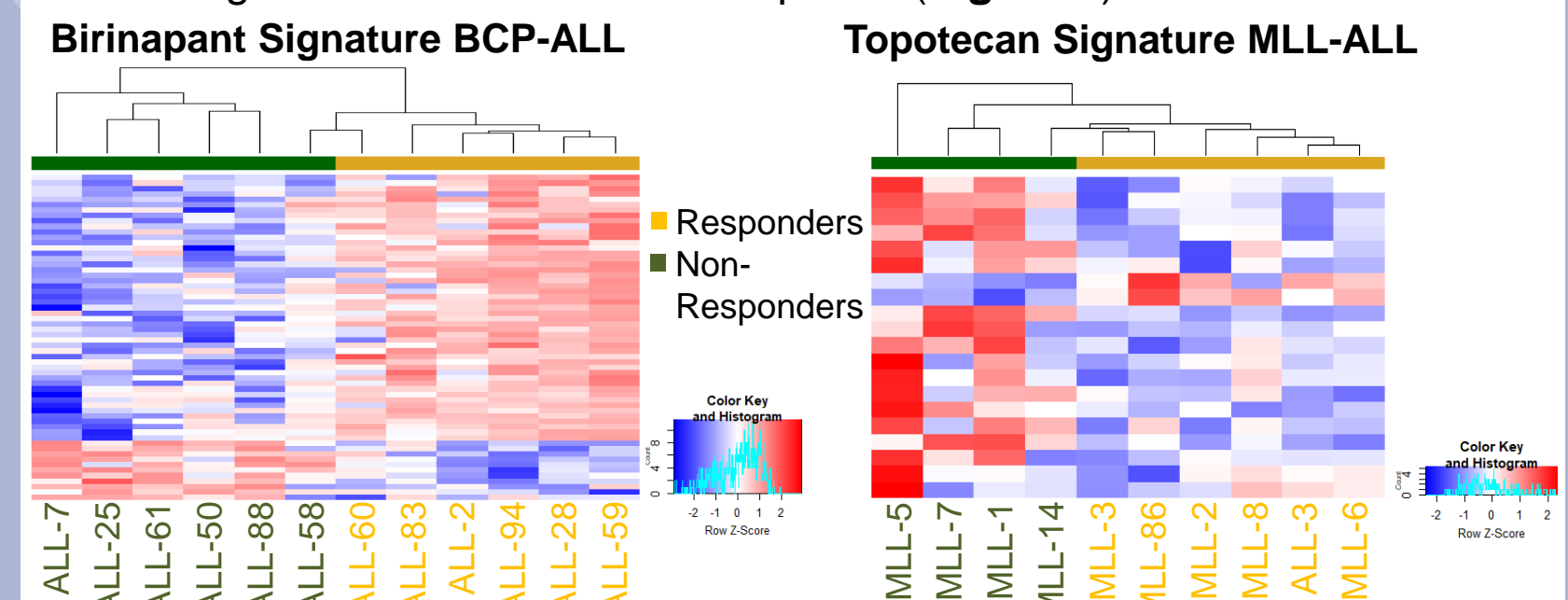


Figure 4. Gene expression signatures associated with birinapant and topotecan sensitivity in BCP-ALL and MLL-ALL, respectively. (Left) A 61 gene signature (rows) separated the basal gene expression profiles of 12 BCP-ALL PDXs (columns) based on their *in vivo* response to birinapant. (Right) A 20 gene signature (rows) separated the basal gene expression profiles of 10 MLL-ALL PDXs (columns) based on their *in vivo* response to topotecan.

4. Discussion and Conclusions

- SMTs provide an accurate and cost-effective platform for preclinical testing of novel agents on an almost clinical trial scale.
- With the elimination of PDX stocks contaminated by thymoma and by using NSG mice instead of the historically used NOD/SCID strain (which has a propensity to develop thymoma), the success rate of SMTs is predicted to be > 90%.
- SMTs can almost encompass the heterogeneity of pediatric ALL in a single experiment.
- SMTs have the power to identify candidate molecular biomarkers or gene expression signatures associated with *in vivo* response to established and novel agents in pediatric ALL, when combined with molecularly annotated PDXs. Further work would be needed to validate these biomarkers.

5. References

- Houghton PJ, et al. The Pediatric Preclinical Testing Program: description of models and early testing results. *Pediatr Blood Cancer*; 2007;**49**:928-40.
- El-Hoss J, et al. A single nucleotide polymorphism genotyping platform for the authentication of patient derived xenografts. *Oncotarget*; 2016;**7**:60475-90.
- Jones L, et al. A review of new agents evaluated against pediatric acute lymphoblastic leukemia by the Pediatric Preclinical Testing Program. *Leukemia*; 2016;**30**:2133-2141.

More Information
 *Corresponding author: Richard B. Lock, PhD Email: rlock@ccia.org.au
 Presented at: American Society of Hematology (December 2018)
 Supported by: U01CA199000 and U01CA199222 from the NCI.
www.ncipptc.org
 There are no relationships to disclose.